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(54) Title: PROCESS FOR THE PRODUCTION OF A POROUS CROSS-LINKED POLYSACCHARIDE GEL AND ITS USE AS A GEL FILTRATION MEDIA AND IN CHROMATOGRAPHY

(57) Abstract

A process for the production of a porous cross-linked polysaccharide gel and a gel obtainable by the following steps: a) preparing a solution or dispersion of the polysaccharide; b) adding a bifunctional cross-linking agent having one active site and one inactive site to the solution or dispersion from step a); c) reacting hydroxylgroups of the polysaccharide with the active site of the cross-linking agent; d) forming a polysaccharide gel; e) activating the inactive site of the cross-linking agent; f) reacting the activated site from step e) with hydroxylgroups of the polysaccharide gel, whereby cross-linking of the gel takes place. The cross-linked polysaccharide gel obtained can further be cross-linked by conventional methods, one or several times.

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**PROCESS FOR THE PRODUCTION OF A POROUS CROSS-LINKED POLYSACCHARIDE GEL
AND ITS USE AS A GEL FILTRATION MEDIA AND IN CHROMATOGRAPHY**

5 The present invention relates to a process for the pro-
duction of a cross-linked polysaccharide gel with improved
qualities and a gel obtainable by the process and use
thereof. More precisely the invention refers to a new
method of cross-linking, in which a bifunctional cross-
10 linking agent is introduced into the polysaccharide solu-
tion before emulsion and gel formation.

Chromatographic methods are commonly used for separa-
tion and purification of molecules such as proteins, nu-
cleic acids, polysaccharides etc. A wide variety of separa-
15 tion media is available, both inorganic material as well as
synthetic polymers and polysaccharides.

Gel matrices of polysaccharides have long been used as
separation media due to their good qualities and such ma-
trices are now standard equipment in biochemistry laborato-
20 ries. The polysaccharides are inert to biomolecules under a
wide range of conditions. The polysaccharides are natural
materials and as such are approved of by authorities (such
as the Food and Drug Administration (FDA) in USA) for many
fields of application. When using chromatographic separa-
25 tion methods, there can be left traces of the separation
medium left in the separated product. When polysaccharides
are used, as separation medium, this is harmless, as the
material is not toxic.

Generally, chromatographic separations are carried out
30 in columns packed with the separation matrix in form of
particulate beads. Separation media of a fast kinetics with
rapid flow rates results in a high productivity and may be
achieved by a reduction of the particle size. However,
small beads result in a higher back pressure due to the
35 narrowing of the convective flow channels between the par-
ticles in a packed bed. To be able to separate large mole-
cules the particles should have large pores, but large pores
may result in a weakened structure of the particles. As the

polysaccharides are soft materials the particles may easily collapse, especially at high flow rates. Thus, there is a demand on methods of manufacturing more stable polysaccharide particles. It is well known to increase the stability of polysaccharide particles by cross-linking the polymer. The cross-linking process stabilises the polysaccharide gel matrices by chemically binding the polymer chains with each other at their respective free hydroxyl groups. The cross-linking takes place between the hydroxyl and the functional groups of the cross-linkers. This affects the particle rigidity, but to a lesser extent or not at all the size of the pores. There are several patents and articles disclosing different cross-linking methods. Well known cross-linking agents are epichlorohydrin, bis-epoxides, divinyl sulphone.

In EP 203 049 it was found that the rigidity of the polysaccharides was considerably improved when the cross-linking agent used was monofunctional but also contained an additional masked functional group that could be activated later. The cross-linking was made in two steps. First the polysaccharide was derivatized with the monofunctional group. Then, in a next step the masked group was activated and made to react with the hydroxyl groups of the polysaccharide. In this manner the length of the cross-linking was controlled and the desired rigidity obtained.

The common characteristic for the state of the art methods is that the cross-linking is made on the polysaccharide polymer after the formation of the gel particles. Thus, the cross-linking is made on the ready made structure. Particles of e.g. agarose are prepared by dissolving the agarose in water by heating. The hot water solution is then emulsified to form spherical particles in an organic solvent such as toluene. The particles are precipitated after cooling. The particles are then cross-linked. By varying the concentration of the agarose solution, different pore sizes can be obtained. The lower the concentration of the agarose solution the larger pores are obtained.

The object of the present invention was to obtain an improved process for the production of a cross-linked polysaccharide gel.

A further object of the invention was to produce rigid
5 polysaccharide gel particles with improved capability to withstand high flow rates/back pressures, but with retained separation qualities.

The objects of the invention are achieved by the process and the polysaccharide gel as claimed in the claims.
10 According to the invention a process for the production of a porous cross-linked polysaccharide gel is obtained, which process is characterized by the following steps:

- a) preparing a solution or dispersion of the polysaccharide,
- 15 b) adding a bifunctional cross-linking agent having one active site and one inactive site to the solution or dispersion from step a),
- c) reacting hydroxyl groups of the polysaccharide with the active site of the cross-linking agent,
- 20 d) forming a polysaccharide gel,
- e) activating the inactive site of the cross-linking agent,
- f) reacting the activated site from step e) with hydroxyl groups of the polysaccharide gel, whereby cross-
25 linking of the gel takes place.

According to a further aspect of the invention a porous cross-linked polysaccharide gel is obtainable by the following steps:

- a) preparing a solution or dispersion of the polysaccharide,
30
- b) adding a bifunctional cross-linking agent having one active site and one inactive site to the solution or dispersion from step a),
- c) reacting hydroxyl groups of the polysaccharide with
35 the active site of the cross-linking agent,
- d) forming a polysaccharide gel,
- e) activating the inactive site of the cross-linking agent,

f) reacting the activated site from step e) with hydroxyl groups of the polysaccharide gel, whereby cross-linking of the gel takes place.

With the present invention it was surprisingly found
5 that gels with increased pressure/flow capacities of more than 300 % could be obtained, compared with known gels. It was possible to manufacture highly rigid gel particles also with small particle diameters (about 10 μm).

According to the new method of the invention the cross-
10 linking agent, is introduced into the polysaccharide solution or dispersion before the gel formation. The cross-linking agent is a bifunctional agent with one active site and one inactive site. When added to the polysaccharide solution or dispersion the active site of the agent is al-
15 lowed to react with the hydroxyl groups of the polysaccharide. Thereby the cross-linking agent is chemically bound to the polymer chains before the gel formation process is started. In this manner an internal cross-linking agent is introduced into the polysaccharide.

20 In the first step of the process a solution or dispersion of the polysaccharide is formed. Solvents or dispersing agents commonly used together with polysaccharides can be used such as acetone, acetonitrile, dimethyl sulphoxide, dimethylformamide, pyridine, sec. and tert. alcohols, such
25 as isopropanol, etc. However, according to a preferred embodiment of the invention an aqueous solution of the polysaccharide is used.

After the introduction of the cross-linking agent a gel is formed of the polysaccharide. If water has not been used
30 as the solvent, the solvent or dispersing agent is then disposed of and the polysaccharide is dissolved in water. The gel is formed by emulsifying the water solution in an organic solvent such as toluene or heptane. Then, the inactive site of the cross-linking agent is activated and re-
35 acted with hydroxyl groups of the polysaccharide, whereby the gel is cross-linked.

The cross-linked gel can be further cross-linked by conventional methods as known by the state of the art. This

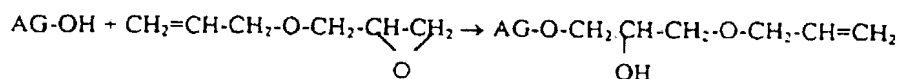
further cross-linking can be made one or several times depending on how rigid particles that are required. The conventional cross-linking can also be made on the gel from step d) before or at the same time as activating and reacting of the inactive site of the cross-linking agent in steps e) and f). In a further embodiment of the invention steps b) and c) can be repeated one or several times after steps d) or e) in order to add more cross-linking agent before performing steps e) and f) or step f).

The bifunctional cross-linking agent used according to the invention comprises one active site and one inactive. With active site is meant all groups capable of reaction with the hydroxyl groups of the polysaccharide. Examples of such groups are halides, epoxides, methanol groups. The inactive site is a group which does not react under the reaction conditions for the reactive site but can later on be activated to react with the hydroxyl groups of the polysaccharide. Groups containing double bonds such as allyl, vinyl, acryl groups are suitable. The group connecting the active and inactive site is not essential, it should however, lack binding activity and not be too long. Preferable cross-linking agents are allylglycidyl ether and allylhalides, preferably allylbromide, but it is also possible to use e.g. N-methylol acrylamide, vinyl benzylchloride, cinnamoyl chloride. The reactions between the hydroxyl groups and the active site and the activated inactive site, as well as the activation of this site, is well known chemistry per se.

The reaction with the bifunctional cross-linking agent could be illustrated on agarose (AG) with the following reaction formulae:

Reaction with the active site of allylglycidyl ether:

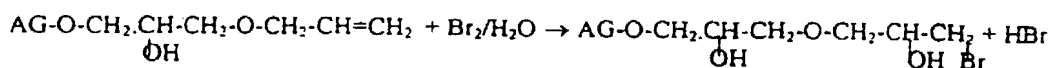
NaOH



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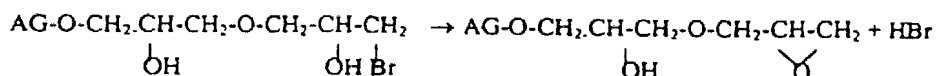
Activation and reaction of inactive site:

pH=7

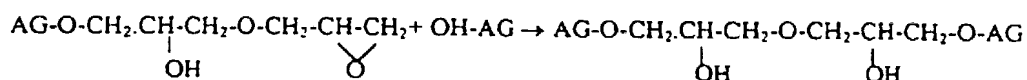


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NaOH



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20 The further cross-linking by conventional methods can be obtained by any of the known cross-linking agents. Suitable compounds are one or several from the group of epihalohydrin, bis-epoxides, divinylsulphone, allylglycidyl ether and dibromopropan-1-ol. Thus, the conventional cross-
25 linking can be made with the same cross-linker as in the internal first cross-linking step or with another cross-linker or with a mixture of cross-linkers.

The process according to the invention can be used on all type of polysaccharides such as agarose, agarose de-
30 rivatives, starch derivatives, dextran or derivatives thereof, alginate, carrageenan. However, agarose is the preferred one.

The gel matrix according to the invention is preferably prepared as particles. The manufacture of the gel is made
35 with well known methods. Agarose for example, is dissolved in water by heating the solution above the gel point for agarose. The cross-linking agent is added to the hot aqueous agarose solution and the active site of the agent is allowed to react with the hydroxyl groups of the agarose.

The agarose solution is then emulsified in an organic solvent such as toluene. The gel particles are precipitated by cooling. Thereafter, the inactive site of the cross-linking agent is activated and reacted with hydroxyl groups of the agarose particles, whereby the gel is cross-linked.

The size of the particles is determined by the stirring speed when emulsifying the agarose solution. The final required particle size is obtained by sieving. The pore sizes are regulated e.g. by varying the polysaccharide concentration. The process according to invention can be used to manufacture polysaccharide particles with conventional diameters and pore sizes. For the production of agarose particles the concentration suitably is from 0.5 - 20 % by weight, preferably from 1 - 12 % by weight. The particle diameters can be from 1 mm - 1 μ m, preferably from 500 μ m - 1 μ m, most preferably 200 μ m - 1 μ m.

With the invention it is possible to produce highly rigid polysaccharide particles. The major parameter that influences the rigidity is the amount of added cross-linker, even if also the polysaccharide concentration has a significance for the rigidity and not only for the pore size as mentioned above. To obtain rigid particles the cross-linker concentration should preferably be within the range 30 - 80 μ mol/g gel, most preferably 45 - 60 μ mol/g gel. A concentration lower than 30 μ mol/g tends to give gels with relatively low pressure/flow capacities. A concentration above 80 μ mol/g can result in gels which shrink too much to be acceptable.

The porous cross-linked polysaccharide gel according to the invention can be used as a gel filtration medium in which the molecules are separated according to differences in their size. They can also be used, after modification, in different types of affinity chromatography. The gel can be substituted with a lot of different groups in per se known manners. Among such groups can be mentioned:

1. Positively charged groups (primary, secondary, tertiary or quaternary amino groups),

2. Negatively charged groups (e.g. carboxy, phosphonic acid, sulphonic acid etc.)
3. Amphoteric groups
4. Groups with specific affinity (e.g. such as between
5 IgG-binding protein (protein A, G, L etc.) and IgG, lectin and carbohydrate, antigen/hapten and antibody, (strept)avidin and biotin,
5. Complementary nucleic acids/oligonucleotides,
6. Groups with pi-electron systems
- 10 7. Chelating groups
8. Hydrophobic groups.

With these groups the matrix can then be used in affinity chromatography, ion exchange chromatography, hydrophobic interaction chromatography, reversed phase chromatography,
15 phy, covalent chromatography etc.

The invention will now be illustrated by the following examples which however are not intended to limit the invention. With parts and percent are meant parts by weight and percent by weight if not stated explicitly.

20 Example 1:

Preparation of agarose solution

An agarose solution is prepared in a batch reactor by adding 7g agarose to 100ml distilled water under stirring for 2h at 95°C.

- 25 After 2h of reaction the solution is cooled to 70°C and 1ml NaOH 45% and 1,67ml allylglycidyl ether (AGE) are added to the agarose slurry. The reaction continues for 2h under stirring at 70°C. The solution is then neutralised with 0,15ml 60% acetic acid and HCl (pH=7-8).

30 **Emulsion media**

The emulsion media is made in an emulsion reactor by adding 5,3g ethyl cellulose (N-50 emulsifier) to 117ml toluene under stirring at 60°C (the dissolving of N-50 in toluene takes approximately 2h).

35 **Transfer of the agarose solution to the emulsion reactor**

The agarose solution is transferred to the emulsion media. The stirring is regulated to 130 rpm. Agarose gel par-

ticles are thereby formed and their size can be controlled by variation of the rotation speed of the stirrer and the addition of extra N-50.

The desired maximal particle size is 150 μm . If the gel particles are too large the rotation speed can be increased up to 220 rpm and extra N-50 can be added. The maximal particle size is controlled by taking samples approximately every 10 min., which are analysed in a microscope with a built-in size graduation.

Once the 150 μm are reached, the solution is cooled down.

The cooling process

The solution is cooled from 60°C to <25°C in approximately 30 min.

Gel washing process

The gel particles are washed under stirring with 1l ethanol 99,5% which is decanted. The gel is then washed on a nutsch with 4x1l ethanol 99,5% and 4x1l distilled water.

Activating of inactive site of allylglycidyl ether and cross-linking of the agarose = Cross-linking no.1

Bromination

10g NaAc (sodium acetate) are added to a reactor containing a solution of 100ml gel and 200ml distilled water under stirring. After 5 min. bromine-water ($\text{Br}_2/\text{H}_2\text{O}$) is added to the solution until a dark yellow colour is obtained and maintained for over 1 min. The reaction continues for approximately 15 min. thereafter sodium formate is added, giving the gel a white colour.

The gel is washed with 3x1l distilled water.

Cross-linking

5g Na_2SO_4 are added to a reactor containing a solution of 100ml brominated gel and 100ml distilled water under stirring. After 15 min. 10ml NaOH 45% and 0,3g NaBH_4 are added to the solution. The reaction continues for 3h and then the temperature of the solution is increased to 40°C and the reaction goes on for 16h.

The gel is washed with distilled water until the pH=7.

Further cross-linking with a conventional method =
Cross-linking no. 2

5 45,3g Na₂SO₄ are added to a reactor containing a solution of 100ml AGE cross-linked gel and 33,3ml distilled water (75% gel slurry) under stirring. The reaction temperature is increased to 50°C and after 2h, 1,33ml 45% NaOH and 0,4g NaBH₄ are added to the solution as well as 9,33ml NaOH
10 45% and 10ml epichlorhydrin (ECH), which are added during a period of 6-8h. The reaction continues over night (ca. 16h). The gel is washed with distilled water and 60% acetic acid is added to obtain a pH=5-6.

The gel is then sieved to the desired particle size intervals (40-100 µm).

Example 2:

In this example particles are prepared, which are cross-linked twice by conventional methods after the cross-linking according to the invention. Thus, particles prepared according to example 1 were cross-linked with Cross-linking no. 2 method:

118g Na₂SO₄ are added under stirring to a solution of 260ml ECH cross-linked gel and 87ml distilled water. The temperature is slowly increased to 50°C and after 2 hours
25 3,5ml NaOH 45% and 0,35g NaBH₄ are added, while 24ml NaOH 45% and 26ml ECH are slowly pumped (6-8 hours) into the solution. The reaction is kept for 16 hours and then the gel is washed with distilled water (and 0,6ml acetic acid) until pH≈5-6.

30 The gel is then sieved and tested.

Example 3:

As a comparable example agarose gel matrices allylated after the emulsion process are prepared. A synthesis method similar to the synthesis method described in example 1 for
35 allylation before emulsion is used. These products have the same allyl concentration as those produced with the newly developed technique. The alternative synthesis method used

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to produce these gels consists of the following steps:

Emulsion

28g agarose are added to 400ml distilled water and heated to 95°C for 2 hours. Thereafter the solution is cooled to 70°C and transferred to a 60°C warm solution containing 470ml toluene and 35g N-50 (emulsifier). After approximately 45min. stirring an average particle size of 150 µm is obtained. The solution is cooled for about 30 min. to 22°C and washed with absolute ethanol (4x2l) and distilled water (4x2l).

Allylation:

33,5g Na₂SO₄ and 1,8g NaBH₄ are added to a solution containing 355ml agarose gel and 106,5ml distilled water. This solution is stirred for 5 min. at a temperature of 30°C and then 25ml AGE and 71ml NaOH 45% are slowly pumped (6-8 hours) to the solution. The stirring continues for 16 hours at the same temperature. The allylated gel is washed with distilled water (3x2l).

Bromination:

36g sodium acetate is added to a solution of 360ml allylated gel and 720ml distilled water. After 5 min. stirring, 146ml Br₂/H₂O is added to the solution and the reaction is run for 15 min. Then 0,5g sodium formate is added and the gel obtained a white colour. The gel is washed with 3x2 gel volume of distilled water.

Cross-linking no. 1:

18g Na₂SO₄ is added to a solution of 360ml brominated gel and 360ml distilled water. The mixture is kept under stirring and after 15 min. 72ml NaOH 45% is pumped (for 30 min.) into the solution together with 1,08g NaBH₄. After 3 hours the temperature is increased to 40°C and the reaction is held for 16 hours. The gel is then washed with distilled water until pH is about 7.

Cross-linking no. 2:

156,4g Na₂SO₄ is added to a solution containing 345ml cross-linked gel and 115ml distilled water. The solution is kept under stirring and slowly heated to 50°C. After 2

hours 4,6ml NaOH 45% and 0,46g NaBH₄ are added to the mixture while 32,2ml NaOH 45% and 34,5ml ECH are pumped for 6-8 hours to the solution. The reaction continues for 16 hours and then the gel is washed with distilled water (and 0,6ml acetic acid) until pH is about 5-6.
The gel is sieved and tested.

Example 4:

Extra allylation of an allylated agarose gel matrix after cross-linking no. 1

During the allylation process of the agarose solution, the bifunctional AGE molecule binds to the agarose polymer chains with its active site leaving its inactive site free. The free site is first brominated and during cross-linking no. 1 it is epoxidized with NaOH, enabling the AGE molecule to bind to a second agarose polymer chain. The alternative synthesis method here aims at coupling more AGE to the polymer chains by repeating the allylation process one more time before starting the first cross-linking as it is explained experimentally in the following steps:

Bromination:

35g sodium acetate is added to a solution of 350ml allylated gel, prepared according to example 1, and 700ml distilled water. The mixture is kept under stirring and after 5 min. 160ml Br₂/H₂O are added. The reaction continues for 15 min. and then 0,5g sodium formate is added. The gel colour changes to white and the gel is washed with distilled water (3x1l).

Allylation:

17,5g Na₂SO₄, 0,5g NaBH₄ and 35ml NaOH 45% are added to a solution of 350ml brominated gel and 175ml H₂O. The mixture is held under stirring 40°C and after 1 hour 20ml AGE are added. The reaction continues for 16 hours after which the gel is washed with distilled water until pH is about 7.

Bromination:

38,5g sodium acetate are added to a stirred solution containing 385ml gel allylated twice and 770ml distilled water. After 5 min. 85ml Br₂/H₂O are added and the reaction is kept for 15 min. and then 0,5g sodium formate are added

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giving the gel a white colour. The gel is then washed with distilled water (3x1l).

Cross-linking no. 1:

19,25g Na_2SO_4 are added to a stirred mixture containing
5 385ml of the above mentioned brominated gel and 385ml dis-
tilled water. After 15 min. 38,5ml NaOH 45% is pumped to
the solution (pumping period = 30 min.) along with 1,16g
NaBH₄. The reaction continues for 45 min. before the tem-
perature is increased to 40°C. The solution is kept under
10 stirring for 16 hours. The gel is then washed with dis-
tilled water until pH is about 7.

Cross-linking no. 2:

174,4g Na_2SO_4 are added to a stirred solution of 385ml
cross-linked gel and 128ml distilled water. The mixture is
15 slowly heated to 50°C and after 2 hours 5,13ml NaOH 45% and
0,51g NaBH₄ are added to the solution while 36ml NaOH 45%
and 38,5ml ECH are slowly pumped (6-8 hours) into the reac-
tor. The reaction continues for 16 hours, and then the gel
is washed with distilled water (and 0,6ml acetic acid) un-
20 til pH=5-6.

The gel is sieved and tested.

Example 5:

Second cross-linking before the first cross-linking

In this synthesis the ECH cross-linking is performed
25 before cross-linking no. 1. The experimental procedure is
explained in the following steps.

Cross-linking no. 2:

An allylated gel is produced according to the method
described in example 1. 158,8g Na_2SO_4 are added to a solu-
30 tion containing 350ml of this allylated gel and 117ml dis-
tilled water. The mixture is stirred and slowly heated to
50°C and after 2 hours 4,67ml NaOH 45% and 0,67g NaBH₄ are
added to the solution while 32,7ml NaOH 45% and 35ml ECH
are slowly pumped (6-8 hours) into the reactor. The reac-
35 tion is held for 16 hours and then the gel is washed with
distilled water until pH is about 5-6.

Bromination:

20,5g sodium acetate are added to a solution prepared from 205ml ECH-cross-linked gel and 410ml distilled water.

- 5 The mixture is stirred and after 5 min. 70ml Br₂/H₂O are added and after 15 min. reaction time 0.5g sodium formate are added and the gel becomes white. The gel is then washed with distilled water (3x1l).

Cross-linking no. 1:

- 10 10,25g Na₂SO₄ are added to a solution containing 205ml brominated gel and 205ml distilled water. The mixture is stirred for 15 min. and then 20,5ml NaOH 45% and 0,61g NaBH₄ are added. After 3 hours reaction time the temperature is increased to 40°C and the solution is kept under
15 stirring for another 16 hours. The gel is then washed with distilled water until pH is about 5-6.

The gel is sieved and tested.

- The gels prepared in the examples were analysed with respect to allyl concentration, the pressure/flow capacities, K_{av} values (K_{av} is an accepted definition of the
20 relative pore size) and particle size distribution accordingly:

Allyl concentration analysis

- The allyl concentration was analysed on a Mettler
25 DL40GP Memo Titrator with 0.1 M AgNO₃ according to standard methods.

The pressure/flow capacities analysis

Instrument: XK-16/40 column

HR 10/30 column

- 30 FPLCdirector™ system control unit (max. pressure/flow output: 26-30 bar or 200-210 ml/min):
a Pharmacia Biotech Controller LCC-501 Plus and
two Pharmacia Biotech Pump P-6000 and PC-unit
with built-in interface.

- 35 **Method:** The testing of pressure/flow resistance of a gel matrix depends on the gel average bead size and the column packing process. The gel beads produced have two average size intervals a) 8-12 µm b) 40-100 µm.

Packing:

a) The column packing process for gel beads with average size 8-12 μm is made in a HR 10/30 column with a series of pressure/time variations in the same manner as for Kav tests (see next paragraph).

b) The column packing process for gel beads with average size 40-100 μm is a free sedimentation process made in a XK 16/40 column with a bed height of 31 ± 1 cm.

Pressure/flow test:

a) Tests on the HR-10/30 column are made by increasing the flow 0,1 ml/min every 5 min and reading the back pressure variation every 5 min.

b) Tests on the XK-16/40 column are made by increasing the flow 1 or 5 ml/min every 5 min and reading the back pressure variation every 5 min.

The Kav-values determination analysis

Instrument: Pharmacia UVM-detector

Pharmacia Biotech FPLCdirector™ system control

2 channel recorder (plotting unit)

Method: The determination of the Kav-values of the gel matrix, results in an estimation of the pore size of the gel beads. The determination is made on the final product (the cross-linked gel). It is performed by graphic interpretation of the retardation time of several proteins, which have been injected into a column containing gel matrix.

Packing:

The final product is packed in a HR 10/30 column under a pressure of 17 bar. The packing solvent used is a solution with the following composition:

60 g HAc + 1 g Tween™ 20 per 1000 ml

To pack the column 30 g gel is dispersed in 30 g packing solvent.

The gel matrix is first packed under a pressure of 6 bars for 50 min then under 17 bars under 10 min.

Protein injection and Kav determination:

The Kav-values are determined for four proteins:

- Thyroglobulin (MW = 669000 g/mol)
- Ferritin (MW = 440000 g/mol)
- BSA (MW = 67000 g/mol)
- R-nase (MW = 13700 g/mol)

5 These proteins are injected into the column one or two at the time (to prevent overlaps to occur).

The eluent media used during this procedure is a buffer solution with pH = 7,2 and the following composition:

- 50 mM Sodium dihydrophosphate ($\text{NaH}_2\text{PO}_4 \times 2\text{H}_2\text{O}$)
- 10 • 150 mM Sodium chloride (NaCl)
- 0,02 % Sodium azide (NaN_3)

To determine the Kav-values it is necessary to know the volume occupied by the void (volume around the agarose beads) which is done by injecting blue dextran into the
15 column.

The obtained plots are interpreted and the data is analysed with a PC which calculates the desired Kav-values.

The particle size distribution analysis:

The mean particle size distribution (d50v) was performed
20 with a Coulter Multisizer.

The results from examples 1 - 5 are compared with a standard agarose gel (Sepharose®6FF) and presented in the following tables:

Table 1a:

Gel no.	Allyl conc. [$\mu\text{mol/g}$ gel]	d50v [μm]	Pressure/Flow test			
			Max flow [cm/h]	Max Pres. [bar]	Pressure Increase	
					>1 bar*	>1,5 bar*
Seph. 6FF	-	91.1	1050	6.00	2.25	3.50
Example 1	53	74.9	3450	17.25	12.00	14.25
Example 2	53	74.9	4500	17.25	12.90	14.10
Example 3	45	71.4	1500	13.00	4.00	5.25
Example 4	46	77.2	$\geq 6000^{**}$	12.90	>12.90	>12.90
Example 5	46	83.7	5250	18.80	13.90	15.10

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* Pressure for which the pressure increase is over 1 or 1.5 bar when the flow is increased by 150 cm/h every 5 min.

** ≥ 6000 cm/h indicates the maximal flow the instrument is capable of delivering. The maximal flow capacity of the gel lies above this value.

Table 1b:

Gel no.	Allyl conc. [$\mu\text{mol/g}$ gel]	Kav			
		Thyro	Ferritin	BSA	R-nase
Seph. 6FF	-	0.37	0.48	0.64	0.81
Example 1	53	0.34	0.48	0.64	0.82
Example 2	53	0.33	0.48	0.64	0.80
Example 3	45	0.28	0.44	0.61	0.80
Example 4	46	0.28	0.43	0.56	0.70
Example 5	46	0.27	0.42	0.58	0.74

10 Example 6:

In this example the method according to the invention was used to prepare highly rigid agarose gel beads with 8.1 w/v % agarose. The process according to example 2 was repeated but when preparing the agarose solution 8.1 g agarose per 100 ml water was used. The gel was sieved and two fractions with two different particle sizes were obtained, Example 6a and 6b. A further particle size, Example 6c was produced according the same manner. The gel beads were tested in the same way as mentioned above. The prepared particles were compared with conventional particles with an agarose content of 8.1 w/v % (Superose®6 from Pharmacia). The test results are put together in tables 2a and 2b.

Table 2a:

Gel no.	Allyl conc. [$\mu\text{mol/g}$ gel]	d50v [μm]	Pressure/Flow test	
			Max flow [ml/min]	Max Pres [bar]
Superose 6	-	13.2	0,9	15
Superose 6	-	14.6	1,2	17
Example 6a	59	9.2	≥ 1.7	$\geq 26^*$
Example 6b	59	12.6	≥ 2.5	$\geq 26^*$
Example 6c	57	11.2	≥ 2.5	$\geq 26^*$

- 5 * 26 bar corresponds to the maximal pressure the instrument is capable of delivering. The maximal pressure and flow values obtained above are expected to be higher.

10 Table 2b:

Gel no.	Allyl conc. [$\mu\text{mol/g}$ gel]	d50v [μm]	Kav			
			Thyro	Ferritin	BSA	R-nase
Superose 6	-	13.2	0.29	0.40	0.54	0.68
Superose 6	-	14.6	0.36	-	0.60	-
Example 6a	59	9.2	0.24	0.37	0.50	0.68
Example 6b	59	12.6	0.26	0.39	0.52	0.68
Example 6c	57	11.2	0.21	0.34	0.47	0.63

- 15 In the following examples gels with different agarose content were produced:

Example 7:

- 20 The method according to example 1 was repeated but with different agarose contents. The result is disclosed in table 3a and 3b below. Example 7a with 7 % agarose is identical with example 1.

Example 8:

In this example different agarose contents were used and the method according to example 5 was used. The result is disclosed in the tables below.

Table 3a:

Gel	Agarose [w/v %]	d50v [μ m]	Pressure/Flow test			
			Max flow [cm/h]	Max Pres. [bar]	Pressure In-crease	
					>1 bar*	>1,5 bar*
Seph.4FF	4	98.6	480	1.8	-	-
Seph.6FF	6	91.1	1050	6.0	2.25	3.5
Example 7a	7	74.9	3450	17.25	12.0	14.25
Example 7b	4	84.0	2250	9.00	5.45	6.60
Example 8a	7	78.4	$\geq 6000^{**}$	19.90	>19.90	>19.90
Example 8b	5	61.7	3150	16.80	9.55	12.30
Example 8c	4	-	2700	9.50	6.25	7.40
Example 8d	3	87.5	1800	4.5	>4.5	>4.5

* Pressure for which the pressure increase is over 1 or 1.5 bar when the flow is increased by 150 cm/h every 5 min.

** ≥ 6000 cm/h indicates the maximal flow the instrument is capable of delivering. The maximal flow capacity of the gel lies above this value.

15 Table 3b:

Gel	Agarose [w/v %]	Kav			
		Thyro	Ferritin	BSA	R-nase
Seph.4FF	4	0.57	0.66	0.76	0.87
Seph.4FF	6	0.37	0.48	0.64	0.81
Example 7a	7	0.34	0.48	0.64	0.82
Example 7b	4	0.53	0.63	0.76	0.87
Example 8a	7	0.26	0.43	0.57	0.73
Example 8b	5	0.36	0.51	0.63	0.77
Example 8c	4	0.56	0.64	0.78	0.88
Example 8d	3	0.67	0.77	0.84	0.93

Conclusion:

From the tables it can be seen that the use of the new cross-linking method according to the invention results in gels capable of withstanding more than three times higher flow than conventional gel particles or particles prepared according to known methods (Example 3), though the gels have similar K_{av} values.

The excellent behaviour of the gels of the invention can also be illustrated as in figures 1 - 3. In the figures:

Figure 1 is a plot of the flow against the back pressure of examples 1 - 5 and comparable compound.

Figure 2 is a plot of the flow against the back of examples 6a, 6b, 6c and comparable compound.

Figure 3a is a plot of the K_{av} values against the maximal flow for examples 7a, 7b, 8a, 8b, 8c, 8d and comparable compounds.

Figure 3b is a plot of the flow against the back pressure for the same examples as in figure 3a.

From the figures it is evident that for the state of the art particles the back pressure raises quickly when the flow increases above a moderate value, which indicates collapse of the particles if the flow is increased too much. However, the pressure/flow plots for the gels according to the invention show a much lower inclination, indicating that the back pressure only raises slowly when the flow is increased.

In figure 3a the K_{av} values for the gel matrices according to examples 7a,b and 8a,b,c,d has been plotted against the maximal flow. It is readily seen from the diagram that the maximal tolerated flow is increased by 300 % for matrices produced according to example 7 or example 8, independently from the agarose w/v percentage. However, the agarose w/v percentage has an important impact on the gel beads pore size, which is expressed by the K_{av} values, as they increase with the reducing amount of agarose in the gel beads.

Claims

1. A process for the production of a porous cross-linked polysaccharide gel, characterized by the following steps:

- 5 a) preparing a solution or dispersion of the polysaccharide,
- b) adding a bifunctional cross-linking agent having one active site and one inactive site to the solution or dispersion from step a),
- 10 c) reacting hydroxylgroups of the polysaccharide with the active site of the cross-linking agent,
- d) forming a polysaccharide gel,
- e) activating the inactive site of the cross-linking agent,
- 15 f) reacting the activated site from step e) with hydroxylgroups of the polysaccharide gel, whereby cross-linking of the gel takes place.

2. A process according to claim 1, characterized in that the cross-linked polysaccharide gel obtained is
20 further cross-linked by conventional methods, one or several times.

3. A process according to claim 1 or 2, characterized in that the gel from step d) is cross-linked by conventional methods before performing steps e) and f).

25 4. A process according to claims 1 or 2, characterized in that the gel from step d) is cross-linked by conventional methods at the same time as performing steps e) and f).

30 5. A process according to claim 1 or 2, characterized in that steps b) and c) are repeated one or several times after steps d) or e) before performing steps e) and f) or step f).

6. A process according to any of the preceding claims, characterized in that an aqueous solution is prepared of
35 the polysaccharide in step a).

7. A process according to claim 6, characterized in that the gel is formed by emulsifying the aqueous solution from step c) to particles, in an organic solvent.

8. A process according to any of the preceding claims, characterized in that the bifunctional cross-linking agent is allylglycidyl ether or allylbromide.

9. A process according to claim 2-5, characterized in that the further cross-linking by conventional methods is obtained by one or several compounds from any of epihalohydrin, bis-epoxides, divinylsulphon, allylglycidyl ether and dibromopropan-1-ol.

10. A process according to any of the preceding claims, characterized in that polysaccharide is agarose.

11. A porous, cross-linked polysaccharide gel obtainable by the following steps:

- a) preparing a solution or dispersion of the polysaccharide,
- 15 b) adding a bifunctional cross-linking agent having one active site and one inactive site to the solution or dispersion from step a),
- c) reacting hydroxylgroups of the polysaccharide with the active site of the cross-linking agent,
- 20 d) forming a polysaccharide gel,
- e) activating the inactive site of the cross-linking agent,
- f) reacting the activated site from step e) with hydroxylgroups of the polysaccharide gel, whereby cross-linking of the gel takes place.

12. A polysaccharide gel according to claim 11, characterized in that the cross-linked polysaccharide gel obtained is further cross-linked by conventional methods, one or several times.

13. A polysaccharide gel according to claim 11 or 12, characterized in that the gel from step d) is cross-linked by conventional methods before performing steps e) and f).

14. A polysaccharide gel according to claim 11 or 12, characterized in that the gel from step d) is cross-linked by conventional methods at the same time as performing steps e) and f).

15. A polysaccharide gel according to claim 11 or 12, characterized in that steps b) and c) are repeated one or

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several times after steps d) or e) before performing steps e) and f) or step f).

16. A polysaccharide gel according to any of the preceding claims, characterized in that an aqueous solution
5 is prepared of the polysaccharide in step a).

17. A polysaccharide gel according to claim 16, characterized in that the gel is formed by emulsifying the aqueous solution from step c) to particles, in an organic solvent.

10 18. A polysaccharide gel according to any of the preceding claims, characterized in that the bifunctional cross-linking agent is allylglycidyl ether or allylhalide.

19. A polysaccharide gel according to claim 12,
15 characterized in that the cross-linked polysaccharide gel obtained is further cross-linked, by one or several compounds from any of epihalohydrin, bis-epoxides, divinylsulphon, allylglycidyl ether and dibromopropan-1-ol.

20 20. A polysaccharide gel according to any of the preceding claims, characterized in that polysaccharide is agarose.

21. Use of a porous, cross-linked polysaccharide gel according to any of the preceding claims as a gel filtration medium, in affinity chromatography, ion exchange chromatography, hydrophobic interaction chromatography,
25 reversed phase chromatography chelate chromatography, covalent chromatography.

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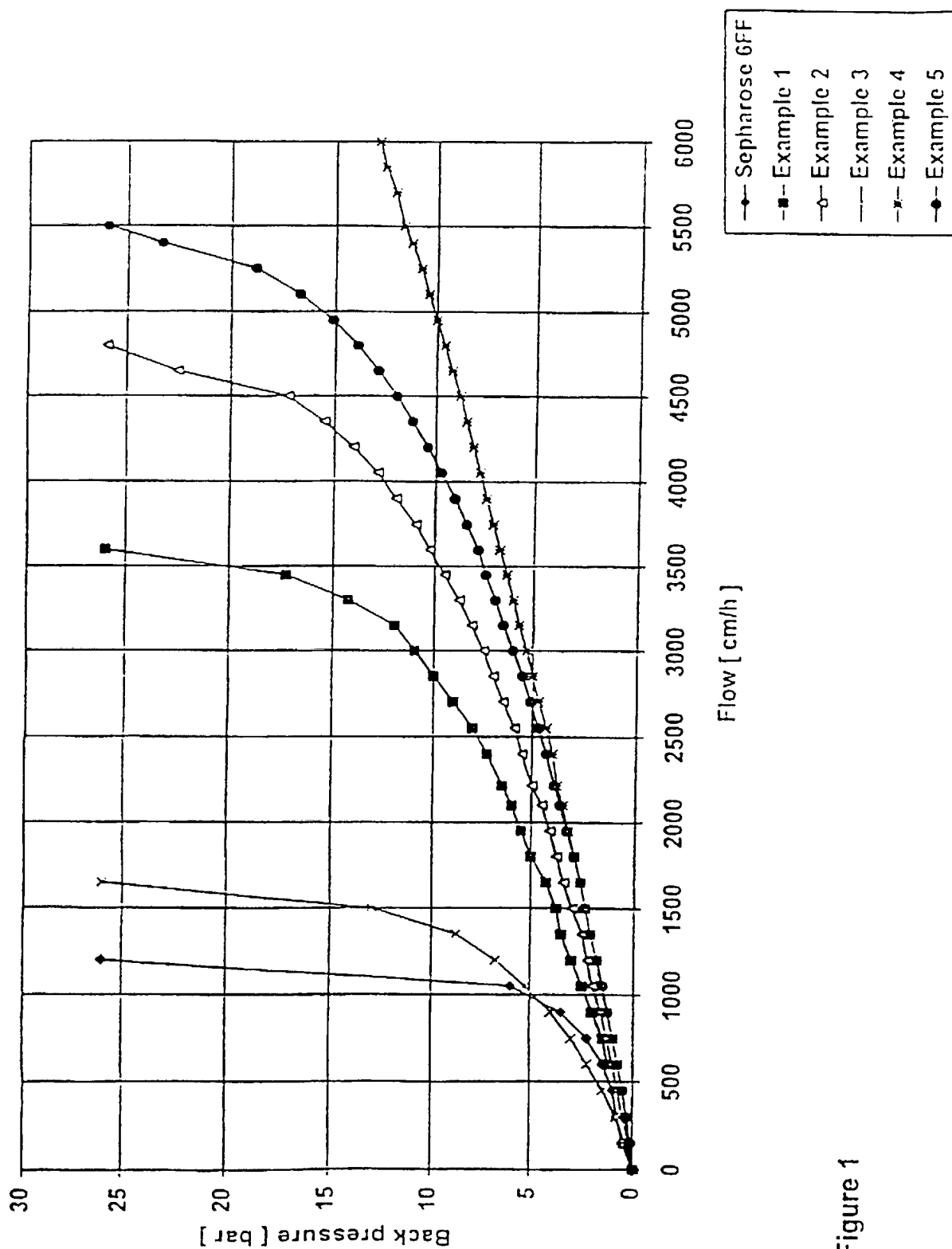


Figure 1

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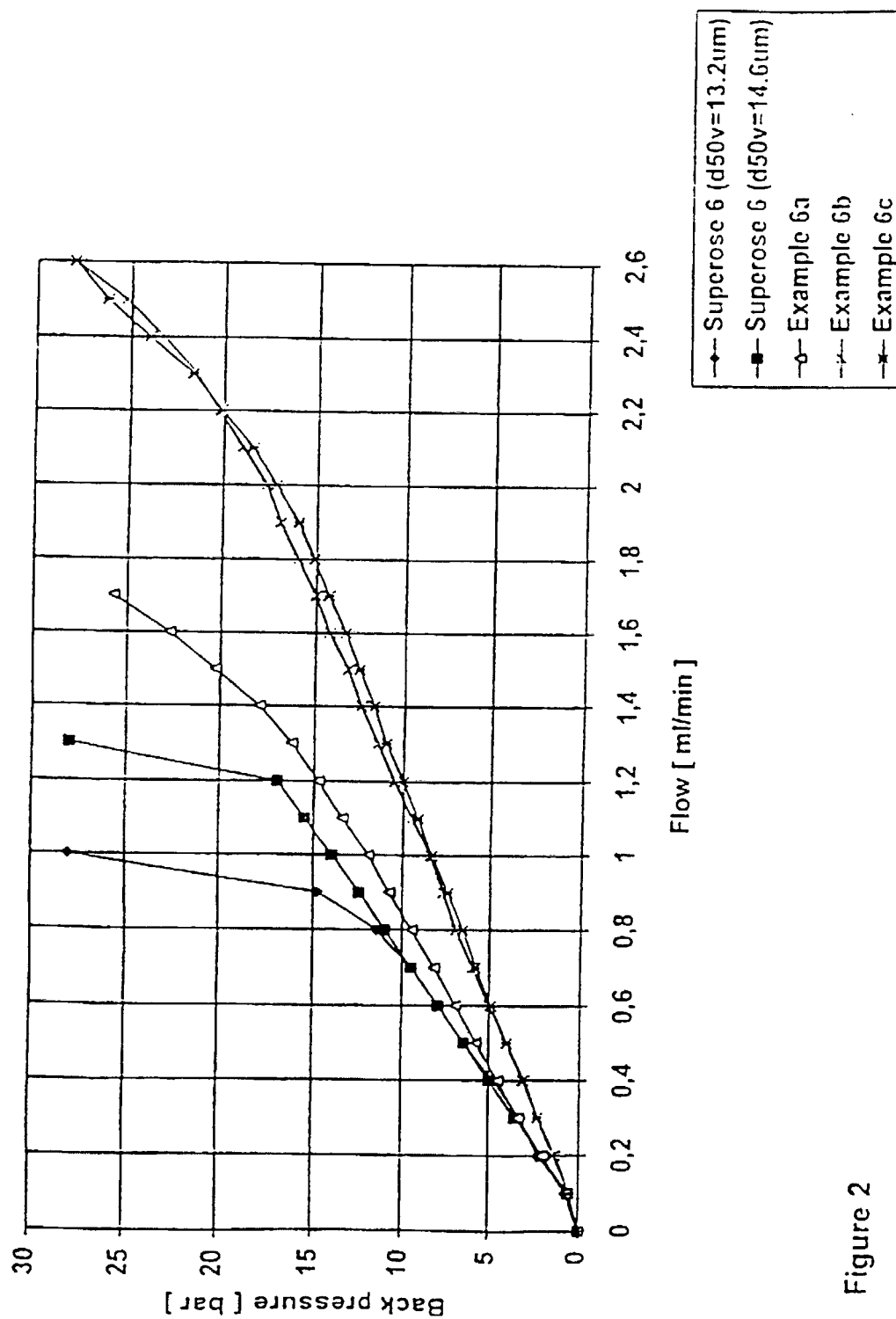


Figure 2

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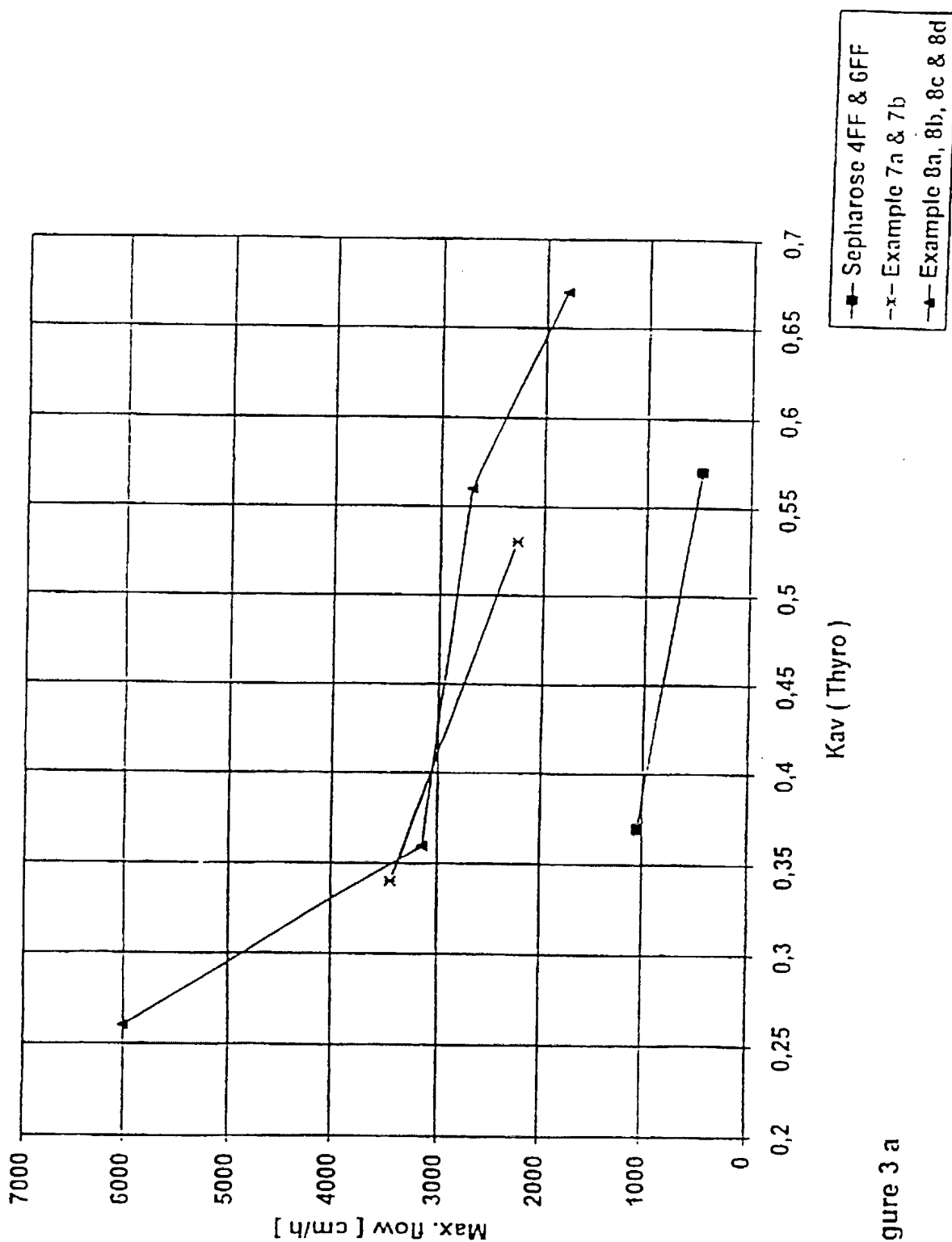


Figure 3 a

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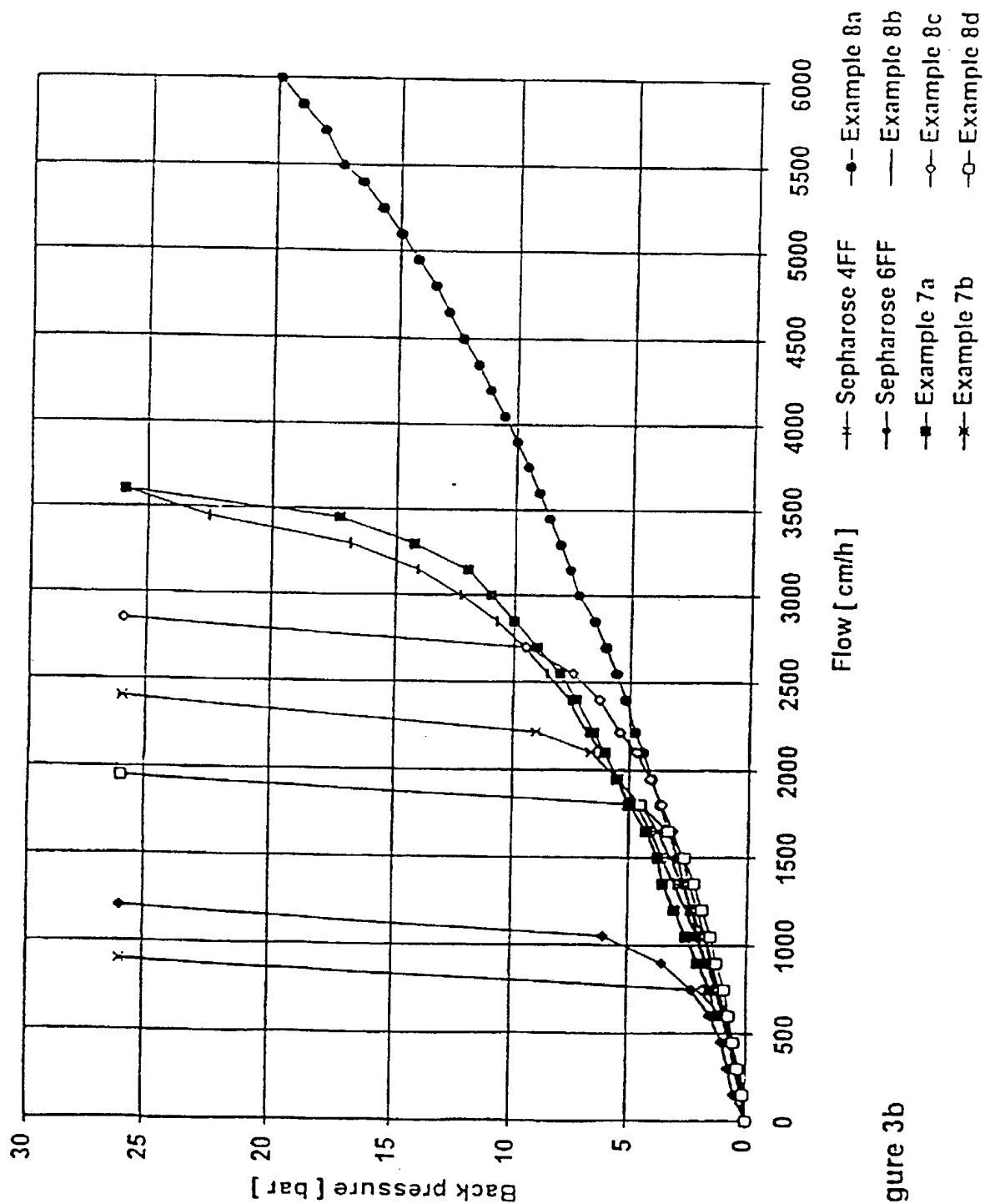


Figure 3b

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 97/00578

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C08B 37/00, C08B 37/12, B01D 15/08

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C08B

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

WPI

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP 0203049 A1 (LKB-PRODUKTER AB), 26 November 1986 (26.11.86), page 3, line 1 - line 10, abstract --	1-21
A	EP 0132244 A1 (PHARMACIA AKTIEBOLAG), 23 January 1985 (23.01.85), page 2, line 14 - line 24, abstract --	1-21
A	US 3860573 A (ERKKI JUHANI HONKANEN ET AL), 14 January 1975 (14.01.75), abstract -- -----	1-21

☐ Further documents are listed in the continuation of Box C.☒ See patent family annex.

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Date of the actual completion of the international search

23 May 1997

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT
Information on patent family members

02/04/97

International application No.
PCT/SE 97/00578

Patent document cited in search report			Publication date	Patent family member(s)	Publication date
EP	0203049	A1	26/11/86	NONE	
EP	0132244	A1	23/01/85	NONE	
US	3860573	A	14/01/75	NONE	

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